

Importance of Microcystins/Nodularins Determination

Most of the world's population relies on surface freshwaters as its primary source for drinking water. The drinking water industry is constantly challenged with surface water contaminants that must be removed to protect human health. Toxic cyanobacterial blooms are an emerging issue worldwide due to increased source water nutrient pollution caused by eutrophication. Microcystins and Nodularins are cyclic toxin peptides. Microcystins (of which there are many structural variants, or congeners) have been found in freshwater throughout the world. To date, approximately 250 variants of Microcystin have been isolated. The most common variant is Microcystin-LR. Other common Microcystin variants include YR, RR, and LW. These toxins are produced by many types of cyanobacteria (blue-green algae), including *Microcystis*, *Anabaena*, *Oscillatoria*, *Nostoc*, *Anabaenopsis*, and terrestrial *Hapalosiphon*. Nodularins are produced by the genus *Nodularia* and they are found in marine and brackish water. Acute poisoning of humans and animals constitutes the most obvious problem from toxic cyanobacterial blooms, and in several cases, has led to death. Human and animal exposure to these toxins occurs most frequently through the ingestion of water, through drinking or during recreational activities in which water is swallowed. These toxins mediate their toxicity by inhibiting liver function and are potent inhibitors of the serine/threonine protein phosphatases, and therefore they may act as tumor promoters.

To protect consumers from adverse health effects caused by these toxins, the World Health Organization (WHO) has proposed a provisional upper limit for Microcystin-LR of 1.0 ppb ($\mu\text{g/L}$) in drinking water. For recreational bathing waters, the WHO has established the following guidelines:

- Relatively low risk of exposure effect at 4 ppb
- Moderate probability of exposure effect at 20 ppb
- High probability of exposure effect – scums

The U.S. Environmental Protection Agency (EPA) has also established guidelines for Microcystins in drinking water:

- For children below school age, 0.3 ppb
- For all other age groups, 1.6 ppb

Performance Data

Test sensitivity: The limit of detection (LOD) for Microcystins and Nodularins with the ABRAXIS® Microcystins Strip Test for Source Drinking Water is 1 ppb. At this level, the test line exhibits moderate intensity. At levels greater than 5 ppb, the test line is not visible. When compared with samples of known Microcystins concentration, it is possible to obtain a semi-quantitative result.

Selectivity: The assay exhibits very good cross-reactivity with all Microcystin cyclic peptide toxin congeners tested to date.

Cell Lysing: When comparing samples lysed using the ABRAXIS® QuikLyse™ reagents and the 3-cycle freeze/ thaw method, average recovery obtained was 114%, SD = 22.7%.

Samples: A sample correlation between the Eurofins Abraxis Strip Test and ELISA methods showed a good correlation.

References

W. J. Fischer, I. Garthwaite, C.O. Miles, K.M. Ross, J.B. Aggen, A.R. Chamberlain, N.A. Towers, and D.R. Dietrich, Congener-Independent Immunoassay for Microcystins and Nodularins. Environ. Sci. Technol. 35, 2002, 4849-4858.
Worldwide Patenting PCT WO 01/18059 A2. U.S. Patent Number 6,967,240. U.S. Patent Number 9,739,777.

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ABRAXIS® Microcystins Strip Test

Immunochromatographic Strip Test for the Detection of
Microcystins and Nodularins in Source Drinking Water

ABRAXIS® QuikLyse™ reagents may be used in a method of U.S. Patent 9,739,777

Product No. 520019 (5 Test), 520020 (20 Test)

1. General Description

The ABRAXIS® Microcystins Strip Test for Source Drinking Water is a rapid immunochromatographic test designed for use in the qualitative screening of Microcystins and Nodularins in source drinking water (please see the Gold Standard Diagnostics Horsham website for information on the screening of brackish or sea water samples and for sample preparation and testing of benthic mat samples). A rapid cell lysis step (ABRAXIS® QuikLyse™) performed prior to testing is required to measure total Microcystins (dissolved, or free, plus cell-bound). The ABRAXIS® Microcystins Strip Test provides only preliminary qualitative test results. If necessary, positive samples can be confirmed by ELISA, HPLC, or other conventional methods.

2. Safety Instructions

Discard samples according to local, state, and federal regulations.

3. Storage and Stability

The ABRAXIS® Microcystins Strip Kit should be stored between 2-30°C. The test strips, test vials, and water samples to be analyzed should be at room temperature before use. Reagents may be used until the last day of the month as indicated by the expiration date on the box.

4. Test Principle

The test is based on the recognition of Microcystins, Nodularins, and their congeners by specific antibodies. The toxin conjugate competes for antibody binding sites with Microcystins/Nodularins that may be present in the water sample. The test device consists of a vial containing specific antibodies for Microcystins and Nodularins labeled with a gold colloid and a membrane strip to which a conjugate of the toxin is attached. A control line, produced by a different antibody/antigen reaction, is also present on the membrane strip. The control line is not influenced by the presence or absence of Microcystins in the water sample and, therefore, should be present in all reactions.

In the absence of toxin in the water sample, the colloidal gold labeled antibody complex moves with the water sample by capillary action to contact the immobilized Microcystins conjugate. An antibody-antigen reaction occurs forming a visible line in the 'test' area. The formation of two visible lines of similar intensity indicates a negative test result, meaning the test did not detect the toxin at or above the cut-off point established for the toxin. If Microcystins are present in the water sample, they compete with the immobilized toxin conjugate in the test area for the antibody binding sites on the colloidal gold labeled complex. If a sufficient amount of toxin is present, it will fill all of the available binding sites, thus preventing attachment of the labeled antibody to the toxin conjugate, therefore preventing the development of a colored line. If a colored line is not visible in the test line region, or if the test line is lighter than the control line, Microcystins are present at a level of concern (>1 ppb). Semi-quantitative results in the test kit range of 0-5 ppb can be obtained by comparing the sample test strip appearance to the appearance of test strips from solutions of known Microcystins concentrations (control solutions). ABRAXIS® Microcystins controls (PN 422011) are available through Gold Standard Diagnostics.

5. Limitations of the ABRAXIS® Microcystins Strip Test, Possible Test Interference

Numerous organic and inorganic compounds commonly found in water samples have been tested and found not to interfere with this test. However, due to the high variability of compounds that might be found in water samples, test interferences caused by matrix effects cannot be completely excluded.

Mistakes in handling the test can also cause errors. Possible sources for such errors include:

Inadequate storage conditions of the test strip, too long or too short incubation times, extreme temperatures during the test performance (lower than 10°C or higher than 30°C).

The test is designed for use with non-chlorinated source drinking water (freshwater). The use of the test with brackish or seawater samples will produce inaccurate results. Please see the Gold Standard Diagnostics Horsham website for additional information on the testing of brackish or seawater samples using the ABRAXIS® Microcystins Strip Test for Finished Drinking Water. The ABRAXIS® Microcystins Strip Test provides only a preliminary qualitative test result. Use another more quantitative analytical method such as ELISA or instrumental analysis to obtain a confirmed quantitative analytical result. Apply good judgement to any test result, particularly when preliminary positive results are observed.

6. Warnings and Precautions

-Use of the ABRAXIS® Microcystins Test Strips without the ABRAXIS® QuikLyse™ reagents will adversely affect the performance of the test, producing inaccurate results. To test samples without using ABRAXIS® QuikLyse™ reagents for cell lysis, such as when testing for free Microcystins only or when testing samples which have been previously lysed (such as those which have undergone the freeze/thaw method), please use the ABRAXIS® Microcystins Strip Test for Finished Drinking Water at 1 ppb, PN 520016 (5 Test) or PN 520017 (20 Test).

-Use only the ABRAXIS® Microcystins test strips and ABRAXIS® QuikLyse™ reagents from one kit lot, as they have been adjusted in combination.

-Test strips, conical test vials, and lysis vials should be kept sealed in their original packaging with desiccant when not in use. Exposure to humidity during storage may adversely impact their performance and give inaccurate results. After initial use in high humidity conditions, remaining kit components should be stored tightly closed with desiccant and refrigerated (2–8 °C) when not in use. Conical test vials and lysis vials stored with indicating desiccant which has turned from blue to pink (indicating excessive exposure to moisture) should not be used and should be discarded.

-All reagents and samples should be allowed to reach room temperature before testing.

-Prior to use, ensure that the product has not expired by verifying that the date of use is prior to the expiration date on the label.

-Avoid cross-contamination of water samples by using a new sample vial and disposable pipette for each sample.

-Use reasonable judgment when interpreting the test results.

-Results should be interpreted within 5-10 minutes after completion of the test.

7. Sample Collection and Handling

-Collect water samples in glass or polyethylene terephthalate (PETG) containers only. The use of other types of plastic containers may result in adsorptive loss of Microcystins, producing inaccurate (falsely low) results.

-Chlorinated source drinking water samples must be preserved (quenched) with sodium thiosulfate at the time of collection to remove residual chlorine. Samples should be quenched with 0.1 mg of sodium thiosulfate per 1 mL of sample. As sodium thiosulfate can interfere with the QuikLyse reagents, quenched chlorinated source drinking water samples should be manually lysed (freeze/thaw method, etc.) then treated with QuikLyse prior to testing. The quenching of residual chlorine is necessary for treated (chlorinated) water samples only; samples which have not been treated with chlorine do not require additional reagents at the time of collection or manual lysis (freeze/thaw method, etc.) before analysis

-Samples can be stored refrigerated for up to 5 days. If samples must be held for greater than 5 days, samples should be stored frozen.

A. Materials Provided

1. Microcystins test strips in a desiccated container
2. Sample collection vials
3. Unlabeled lysis vials (white capped)
4. Disposable exact volume transfer pipettes (see package for usage instructions)
5. Graduated disposable pipettes (calibrated at 1 mL)
6. Reagent papers
7. Conical test vials
8. Forceps
9. User's guide
10. Vial labels

B. Additional Materials (not provided with the test)

1. Timer
2. ABRAXIS® Microcystins Check Samples (PN 422011) for the preparation of control solutions which can be analyzed with samples, to obtain semi-quantitative sample results (see Section C, Controls, below).

C. Controls

It is a good laboratory practice to use positive and negative controls to ensure proper test performance. Water samples containing known quantities of Microcystins (positive and negative controls) should be analyzed with each lot of test strips to provide a reference for line intensity to be expected.

D. Test Preparation

-Allow the reagents and water sample to reach room temperature before use.

-Remove the number of conical test vials required from the package. The remaining conical test vials are stored in the tightly closed container with desiccant.

E. Procedure

1. When analyzing for total Microcystins (dissolved, or free, and cell-bound), which may be present in source drinking water, **a sample lysis is necessary before analysis**. The ABRAXIS® QuikLyse™ reagents provide a rapid option for cell lysis.
2. Using a new graduated disposable pipette for each sample, **draw the sample to the 1 mL line** (graduation mark slightly below bulb) and **add 1 mL of sample to the lysis vial**.

3. **Cap the vial and shake for 2 minutes**, then allow the sample in the vial to **incubate at room temperature for 8 minutes**, to begin the cell lysis.

4. Using the forceps provided, **add 1 reagent paper to the lysis vial**.

5. **Cap the vial and shake for 2 minutes**, then allow the sample in the vial to **incubate at room temperature for 8 minutes**.

6. **Label conical test vials for each sample** to be tested.

7. Using a new disposable exact volume transfer pipette for each sample, **transfer 200 µL of the previously lysed water sample (Steps 1-4 above) to the appropriately labeled conical test vial** (see pipette package for usage instructions).

8. **Close the conical test vial and shake for 30 seconds**. Examine the vial to ensure all dried reagents are completely dissolved (dried reagents will dissolve, turning the sample purple).

9. **Incubate the conical test vial at room temperature for 20 minutes**.

10. **Insert test strip** (arrows down) into the conical vial.

11. Allow the test to **develop for 10 minutes**.

12. **Remove the test strip. Lay the strip flat and allow to continue developing for 5 minutes**.

13. Read the results visually, as explained below in Section F, Interpretation of Results.

F. Interpretation of Results

Sample concentrations are determined by comparison of the intensity of the test line to the intensity of the control line on the same test strip. Although control line intensity may vary, a visible control line must be present for results to be considered valid. Test strips with a test line which is darker than or of equal intensity to the control line indicates a result which is below the limit of detection of the test. Test strips with a test line which is lighter than the control line indicates a result which is ≥ 1 ppb. Test strips with no test line visible (only the control line is visible) indicates a result which is ≥ 5 ppb. Results should be determined within 5-10 minutes after completion of the strip test procedure. Determination made using strips which have dried for more or less than the required time may be inaccurate, as line intensities may vary with drying time.

<u>Control Line</u>	<u>Test Line</u>	<u>Interpretation</u>
No control line present	No test line present	Invalid result
Control line present	No test line present	≥ 5 ppb
Control line present	Test line present	Between 0 and <5 ppb

The appearance of test strips may also be compared to the illustration below to determine approximate sample concentration ranges. Please note that the illustration is intended for the demonstration of test line to control line intensity only. Results should not be determined by comparing the intensity of test lines from test strips to the test line intensity of the illustration, as the overall intensity of test strips may vary slightly with different lots of reagents. To obtain semi-quantitative results in the test kit range of 0-5 ppb, solutions of known Microcystins concentration (control solutions) must be tested concurrently with samples. Sample test line intensities can then be compared with control solution test line intensities, yielding approximate sample concentrations. Do not use strips run previously to determine semi-quantitative sample concentrations, as test line intensities may vary once strips are completely dry.

Visual Interpretation:



Alternately, test strips can also be interpreted using the AbraScan test strip reader (PN 475025), which provides objective determination of line intensities for consistent interpretation of results as well as a digital photographic record of all test strips.

G. Additional Analysis

If necessary, positive samples can be confirmed by ELISA, HPLC, or other conventional methods. These services are available from commercial analytical laboratories such as GreenWater Laboratories (www.greenwaterlab.com).