

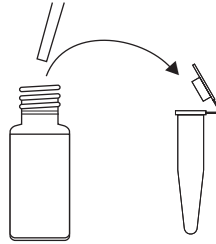
ABRAXIS® Affinity Capture & Extraction (ACE) Kit - Microcystins PN 520100

Prepare Samples

Urine Samples

1. Centrifuge Samples

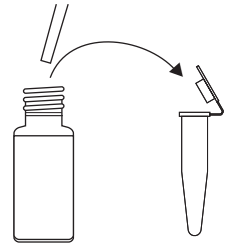
Thaw frozen sample immediately before testing. Add 1.4 mL of thawed sample to a Protein LoBind tube. Centrifuge samples at 2,000 x g for 5 minutes to pellet any precipitates.



Blood Serum Samples

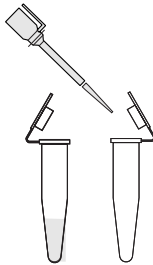
1. Centrifuge Samples

Thaw frozen sample immediately before testing. Add 1.4 mL of thawed sample to a Protein LoBind tube. Centrifuge samples at 10,000 x g for 5 minutes to separate any precipitates or flocculants.



2. Transfer Samples

Transfer the sample supernatant to a new Protein LoBind tube and discard tube with pellet.

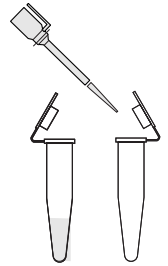


2. Transfer & Dilute Samples

Transfer 0.5 mL of the sample supernatant (clear liquid portion) to a new Protein LoBind tube and discard tube with precipitate.

Add 1.0 mL of Seri-Standard Sample Diluent/Zero Standard, vortex well (1:3 dilution).

NOTE: Beads must be able to move through serum easily and be visible against the magnet. If diluted serum is still too thick to easily pipet, additional Seri-Standard Sample Diluent/Zero Standard may be added. Account for any additional dilutions in final calculation.



3. Run Affinity Capture & Extraction (ACE) Kit

The samples are now ready to be run along with the standards in the ACE Kit.

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Please Note: Prior to running the Affinity Capture & Extraction (ACE) Kit, please prepare the standards, using the flowchart provided in the Standard Set and samples using the flowchart on the backside of this flowchart.

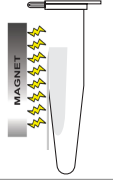
1. Re-suspend Beads

Re-suspend beads with gentle shaking or rotating.



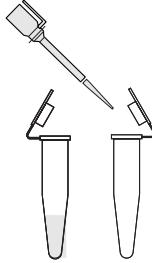
8. Separate Beads from Solution

Pulse spin tubes in microcentrifuge then place tubes in magnetic separator for 2-3 minutes. Discard the supernatant.



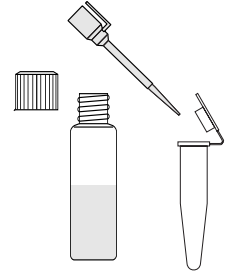
2. Addition of Beads

Add 25 µL of the bead solution for each standard and sample to a Protein LoBind tube. Pool according to user guide instructions.



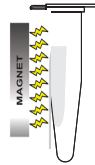
9. Wash Beads

Add 500 µL of the Sample Wash Buffer to the bead solution and vortex.



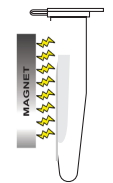
3. Separate Beads from Solution

Place tubes into magnetic separator for 1 minute then discard supernatant



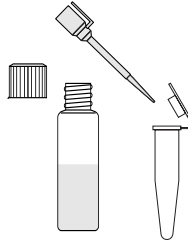
10. Separate Beads from Solution

Pulse spin tubes in microcentrifuge then place tubes in magnetic separator for 2-3 minutes. Discard the supernatant.



4. Wash Beads

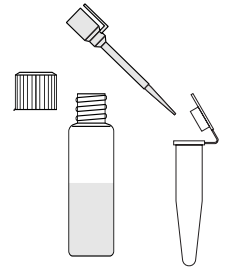
Add 75 µL of the Bead Wash Buffer to each standard and sample and vortex.



11. Add Elution Buffer/Sample Wash Buffer

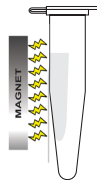
Add either elution buffer or sample wash buffer to the tubes and vortex gently. (For which, please see user guide)

Place tubes into a mixer or thermomixer at 1400 rpm for 5 minutes at the correct temperature.



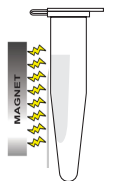
5. Separate Beads from Solution

Place tubes into magnetic separator for 1 minute then discard supernatant



12. Separate Beads from Solution

Pulse spin tubes in microcentrifuge tube then place tubes in magnetic separator for 2-3 minutes.



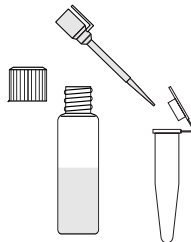
6. Re-suspend Beads

Add 25 µL of the Bead Wash Buffer to each standard and sample and vortex. Separate pool into 25 µL per Protein LoBind Tube.



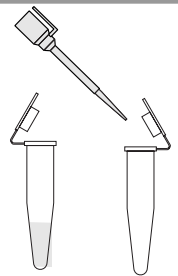
7. Addition of Standards and Samples

Add 1 mL of the standards and samples (previously prepared) to the bead solution and vortex. Place tubes into mixer at 1400 rpm for 5 minutes at room temperature



13. Transfer Elution Supernatant

Transfer the elution supernatant to new Protein LoBind tube and discard beads. Please see user guide for additional post-elution modifications.



14. Run Down Stream Assay

Use the elution supernatant to run the downstream assay.

To run the downstream assay, please refer to the user guide/flow chart provided with the assay. DISCARD the standards that are provided within the downstream assay and replace them with the extracted standards from either the ABRAXIS® Uri-Standards Set Microcystins or the ABRAXIS® Seri-Standards Set Microcystins.