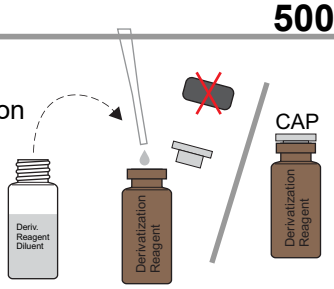
GOLD STANDARD DIAGNOSTICS

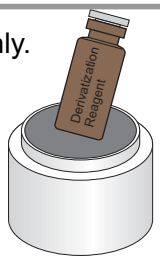
ABRAXIS® Patulin Sample Prep and Derivatization Procedure 500106 & 500110

1. Add

Add 0.5mL of Derivatization Reagent Diluent to Derivatization Reagent.

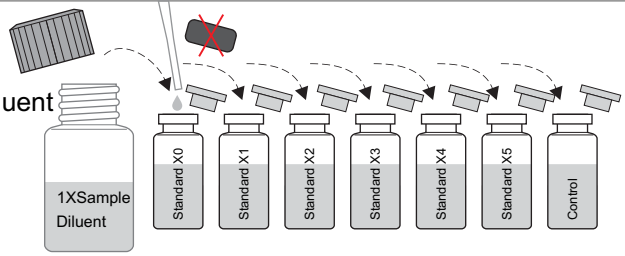


2. Vortex thoroughly.



3. Add

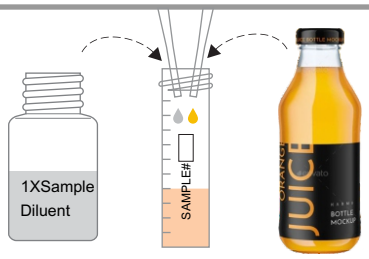
Add 1mL of 1X Sample Diluent to each standard to reconstitute.



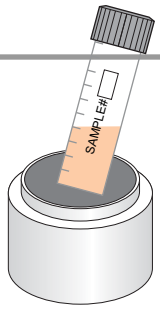
4. Vortex each standard thoroughly.

5. Add

For apple/orange juice, add 4.5 mL 1X Sample Diluent to a labeled tube. Then add 0.5mL sample.

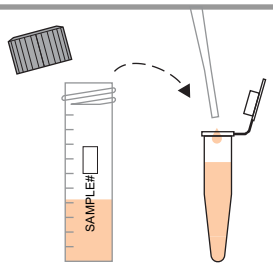


6. Vortex thoroughly.



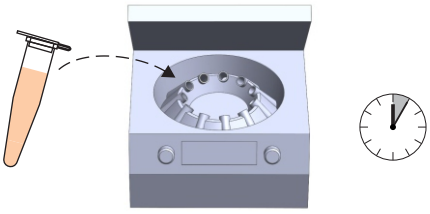
7. Add

Add 2mL of diluted sample to 2mL microcentrifuge tube.



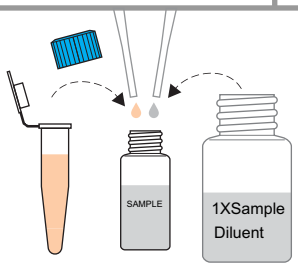
8. Centrifuge

With a balanced centrifuge, spin for 5 minutes at 8000g or 10k rpm.

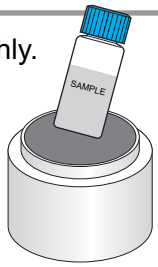


9. Add

Remove 40uL of supernatant and add to 960uL 1X Sample Diluent in a labeled tube.

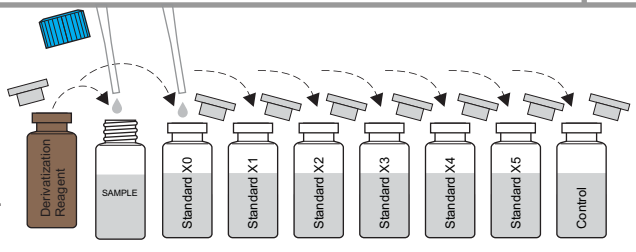


10. Vortex thoroughly.

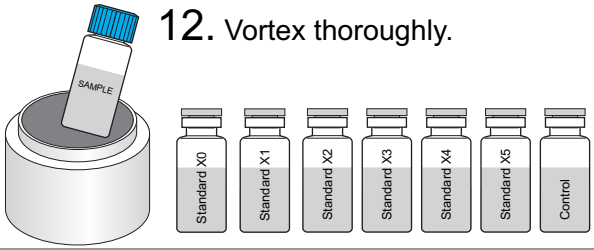


11. Add

Add 10uL Derivatization Reagent to each sample and standard.

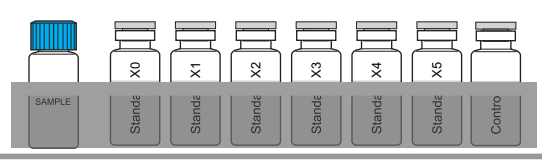


12. Vortex thoroughly.



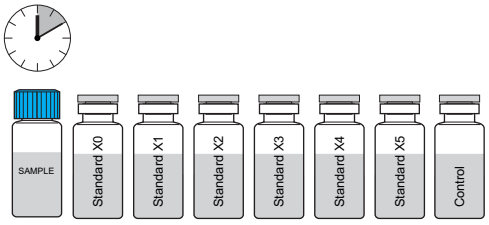
13. Incubate

Incubate standards and samples at 45°C for 45 minutes in a heat block.



14. Cool

After incubation, let the standards and samples cool for at least 10 minutes at RT.



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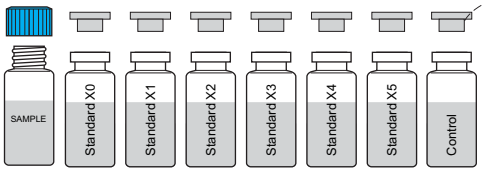
Date this Flow Chart is effective: 05/17/2024

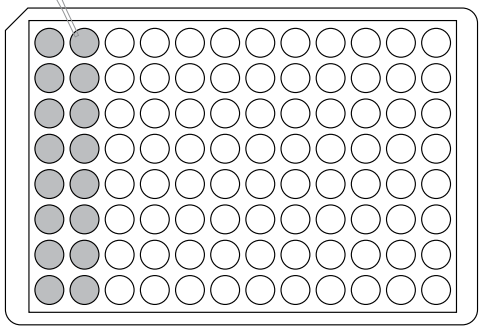
Version: 01

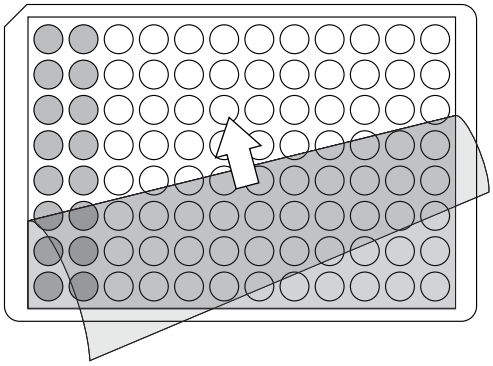
ABRAXIS® Patulin ELISA Plate
500106 & 500110

1. Add

Add 100 uL of the **derivatized** standards, control solutions, samples in duplicate into the wells of the test strips according to the working scheme given.



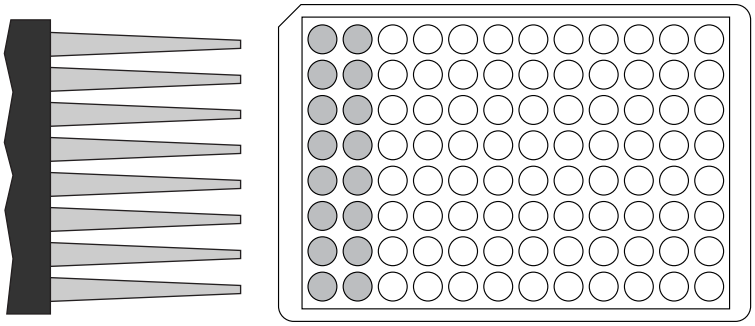


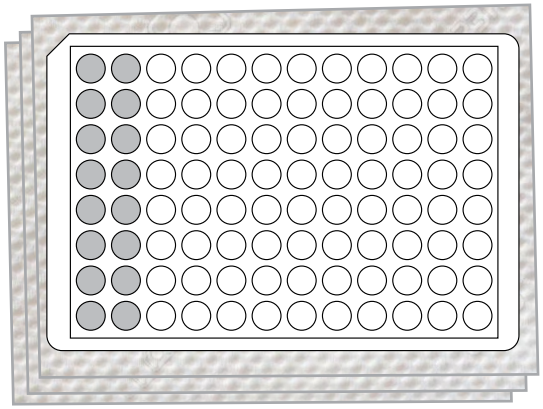


Cover and incubate the strips for 60 min. at room temperature.

2. Wash

After incubation, remove the covering and discard the contents of the wells into a sink. Wash the strips three times (3x) with a multi-channel pipette or repeater pipette using the diluted 1X washing buffer solution (250 uL of washing buffer for each well and each washing step).

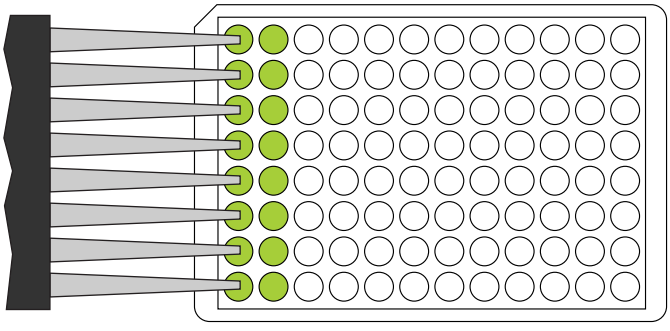




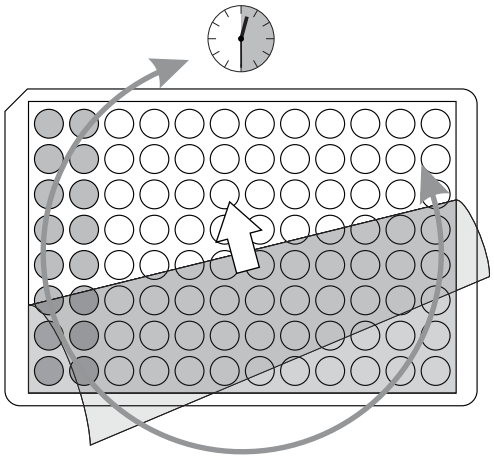
Remaining buffer in the wells should be removed by patting the plate dry on a stack of paper towels.

3. Add

Add 100 uL of the HRP conjugate solution to the individual wells successively using a multi-channel pipette or repeater pipette.



Cover the wells with parafilm or tape and mix the contents by moving the strip holder in a rapid circular motion on the benchtop. Be careful not to spill contents. Incubate the strips for 30 min. at room temperature.



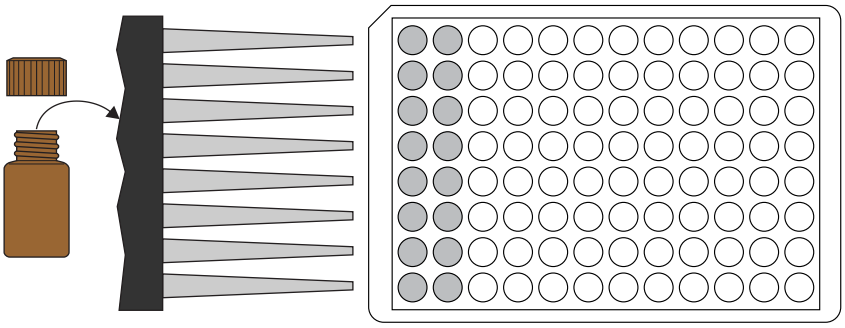
4. Wash (repeat Step 2.)

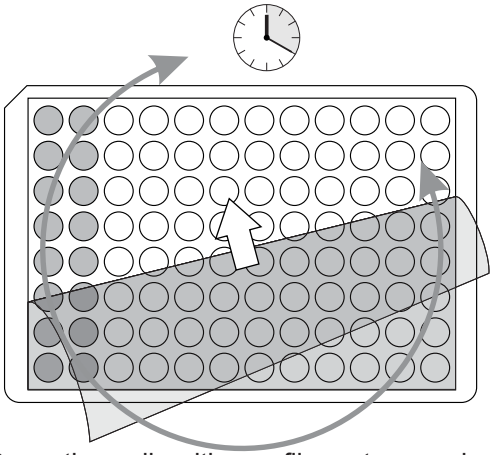
After incubation, remove the covering and discard the contents of the wells into a sink. Wash the strips three times (3x) with a multi-channel pipette or repeater pipette using the diluted 1X washing buffer solution (250 uL of washing buffer for each well and each washing step).

Remaining buffer in the wells should be removed by patting the plate dry on a stack of paper towels.

5. Add

Add 100 uL of substrate/color solution to the individual wells successively using a multi-channel pipette or a repeater pipette.

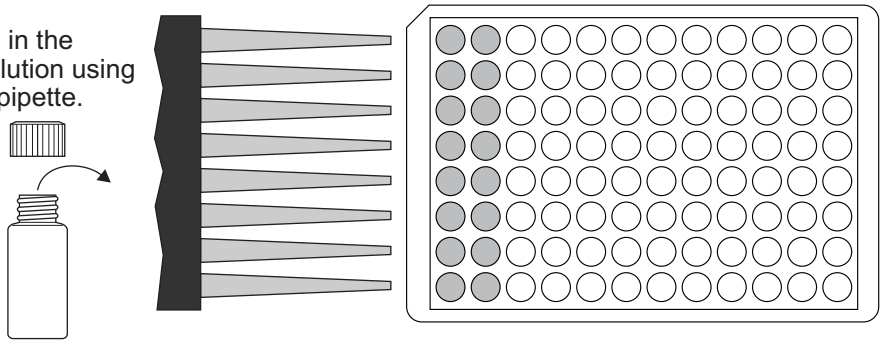




Cover the wells with parafilm or tape and mix the contents by moving the strip holder in a rapid circular motion on the benchtop. Be careful not to spill contents. Incubate the strips for 20 min. at room temperature.

6. Add

Add 100 uL of stop solution to the wells in the same sequence as for the substrate solution using a multi- channel pipette or a repeating pipette.



7. Read

Read the absorbance at 450nm using a microplate ELISA reader. Calculate results.